Plant roots and practical value of plant root symbionts

Conference at Aleksandro Stulginskio University, Akademija 2 May 2016 "Plant Roots: Biology, Morphology, and Functions"

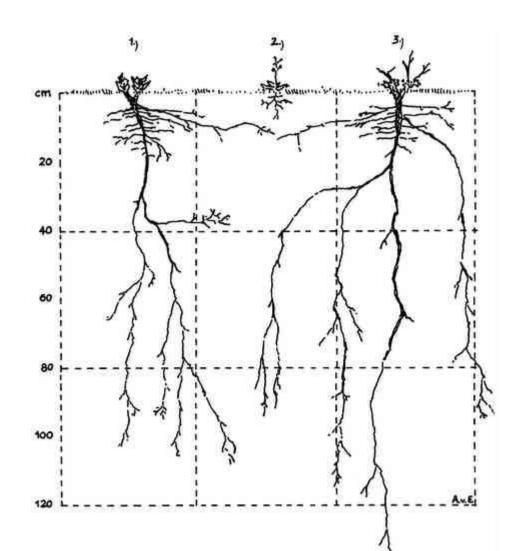
Priv. Doz. Dr. habil Ewald Sieverding University of Hohenheim Institut for Plant Production and Agroecology in the Tropics and Subtropics Garbenstraße 13, Stuttgart-Hohenheim,Germany

> D-55578 Sankt Johann, Auf dem Ewiger 15, Germany sieverdinge@aol.com

What do plant roots do

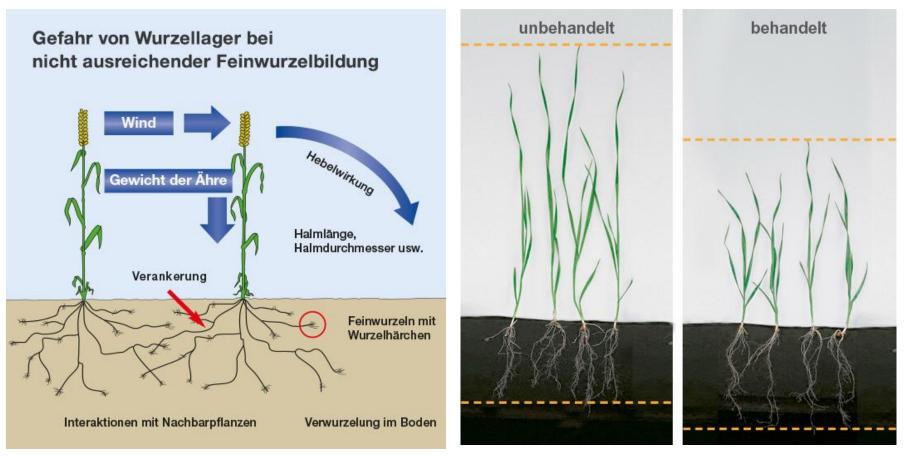
- Fix plants to soil
- Absorb water and nutrients from soil
- Transport water and nutrients to upper parts
- Store assimilates of upper parts (which are remobilized at certain times, or are harvested)
- Exudate components to soil for mineralisation of nutrients
- Act as hosts for symbionts (Rhizobia & Mycorrhiza) and on surface for associations with bacteria and fungi

Roots of different plant species can have very different morphologies



Different root morphologies
1) Aufrechter Ziest; stiff
hedgenettle (*Stachys recta*)
2) Frühes Hungerblümchen;
nailwort (*Erophila verna*)
3) Feld-Beifuß; field mugwort (*Artemisia campestris*)

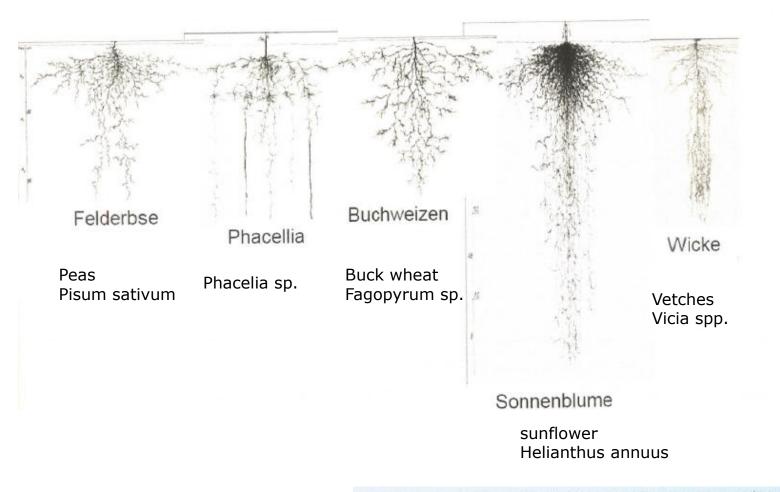
Roots fix plants in soils (cereals) importance of fine roots



Deeper and more roots, and less shoot development give more robust crop

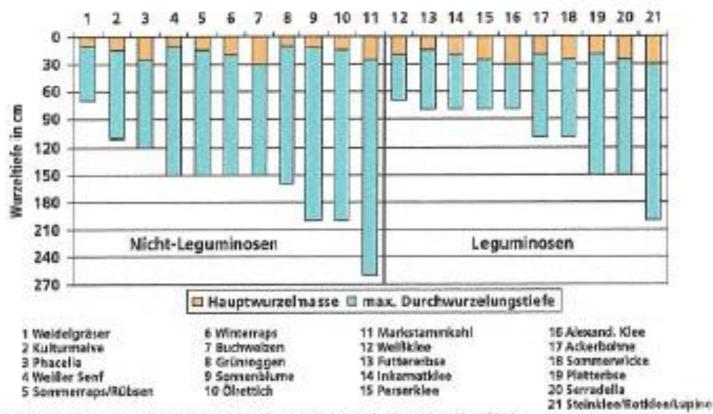
Source: BASF

Roots of crops grow shallow or deep



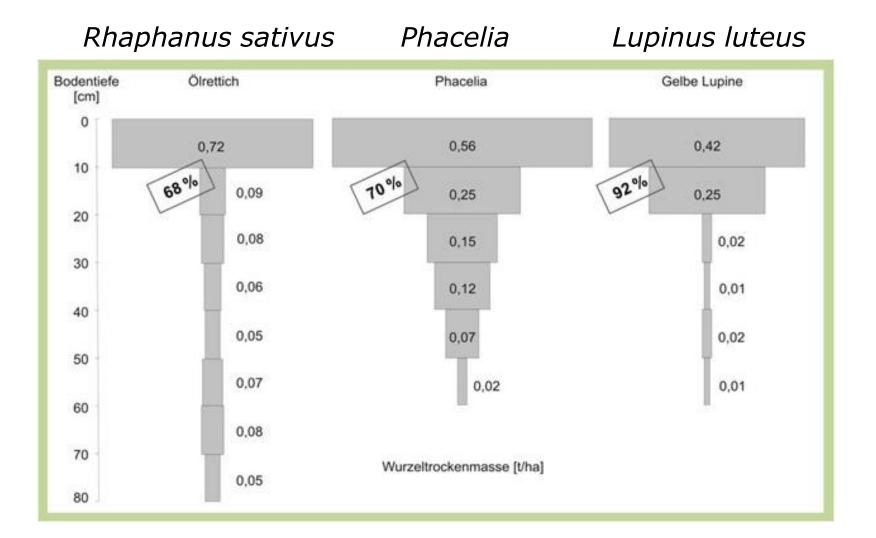
LRA Karlsruhe, Landwirtschaftsamt; Ackerbau-Wasserschutz, Kern

How deep do agronomic plants grow

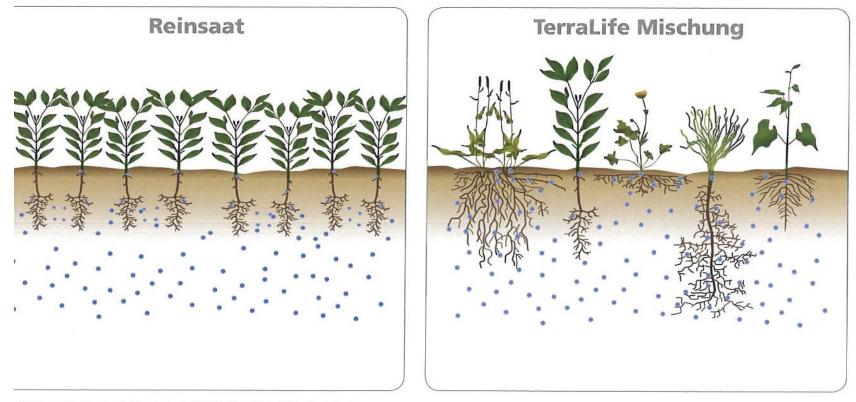


Dualle: Prof. Dr. W. Budiniei, Im Sommer ist Zeit für die Bodersamkrung, Landwirtschaftlichen Wochenblat: 35/2008

Where is most root biomass



Mixed crops can better explore soil for nutrients and water



: DSV, verändert nach Don et. al., 2008 Max Planck Institut, Jena

Mixtures are favoured for intercrops (green manure / greening)

Functional components of roots of some crops

Funktionskomponenten von ausgewählten Zwischenfrüchten

Trockenkeimer	: Bitterlupine, Öllein, Alex., Ramtillkraut, Leindotter,
	Buchweizen, Peluschke, Serradella
Tiefwurzler	: Bitterlupine, Ölrettich (TR), Öllein, Sonnenblume, Alex., So. Wicke, Gelbsenf
Flachwurzler	: Rauhhafer, Ramtillkraut, Buchweizen, Peluschke, Gräser
Schattengarebildner	: Phacelia, Serradella, Ramtilikraut, Sommerwicke,
	Leindotter
N- Sammler	: Bitterlupine, Serradella, Sommerwicka, Peluschke
	Kleearten
Si- Aufschluss	: Öllein
P- Aufschluss	: Buchweizen (anorg. geb. P), Phacelia (org. geb. P)
Allelopathen	: Rauhhafer (Kruziferen, Hirse), Weidelgras (Quecke)
Mykorrhizierer	: Sonnenblume, alle Gräser und Legum., Öllein
Nematodenred.	: Rauhhafer, (Ölrettich), (Senf)
Förder, von Antibios	e: Sommerwicke fördert Bacillus subtiles => bekämpft
T VIGOIL TOIL FUILLETO	Streptomyces scables (Auslöser von K Schorf),
	Rhizoctonia solani und andere
	Tiefwurzier Flachwurzier Schattengarebildner N- Sammler Si- Aufschluss P- Aufschluss Allelopathen Mykorrhizierer Nematodenred.

What influences root development?

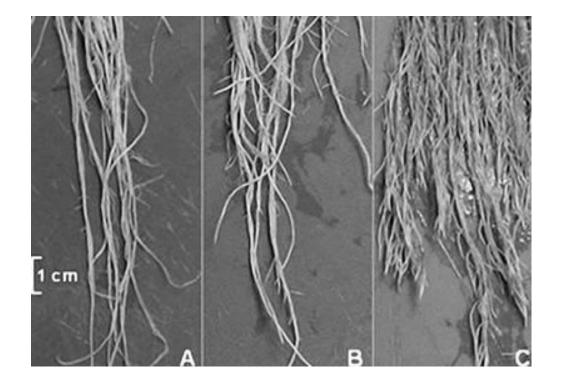


There is are some relationships between availability of nutrients and root development (well known, nothing new)

-The lower the nutrient availability, the better the root formation & vice versa -Phosphate improves root development (- and formation of flowers)

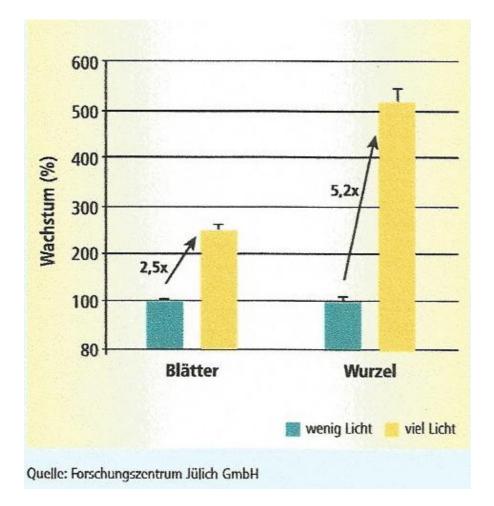
- -Light, temperature and water
- -Airation of soil (compactation)

Phosphate is important for early root development

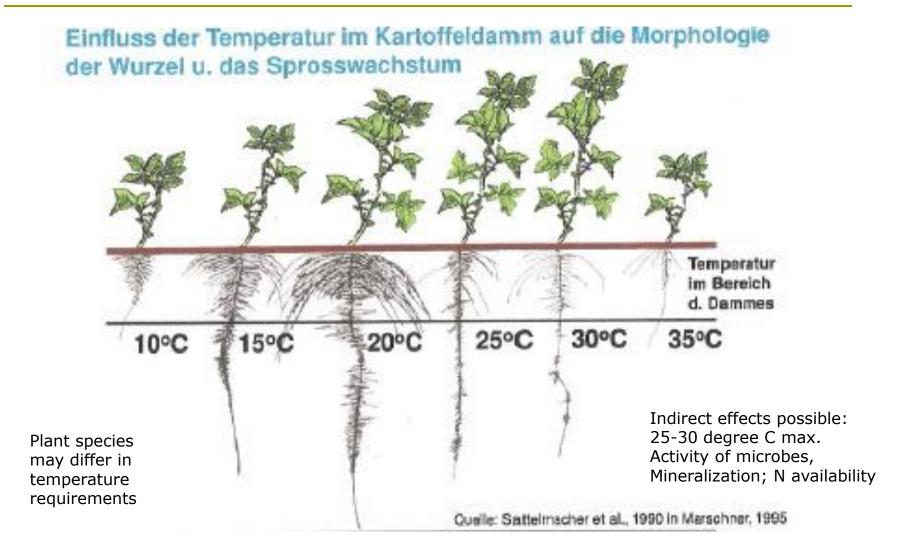


Root development at different P availability (maize)

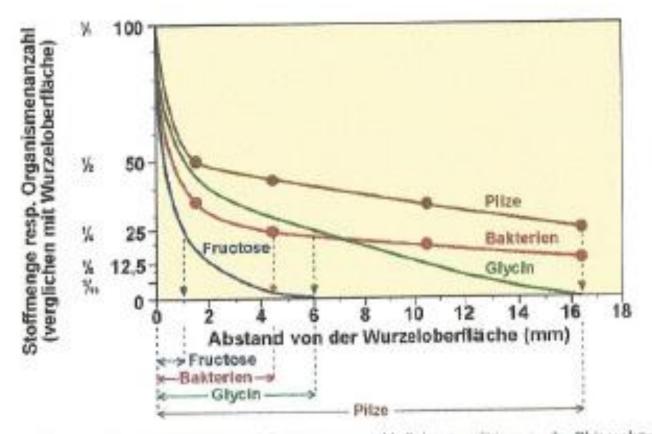
More light – more roots



Influence of temperature



Microorganisms in rhizosphere (living on root exudates)



Stoffmengen (Friedow, Oliveina und Organismenanzahl (Balterior, Pilze) un der Rhizoiphöre. Ausgangsvert an der Wurzeloberfläche = 100% (1/1) Quelle: Gisi,1997

Plant Root Symbionts:

Rhizobia for N2 fixation Mycorrhiza (for nutrient uptake and transport)

Legumes form symbiosis with Rhizobium in roots and fix air nitrogen



Figure 3. Active Nodules on Hoots of Alfalfa (A), Red Clover (C), Fababean (F) and Pea (P)

Source: Microbiology Laboratory, Agriculture Canada Research Station, Switt Current

The Rhizobium-legume symbiosis

soybean plants

Non-inoculated inoculated





soybean root nodules

itrogenase N₂ NH₃

Industrial vs. biological N₂ fixation

Process	10 ⁶ t N year ⁻¹			
Haber-Bosch	100			
Biological N ₂ fixation	>250			

Source: Presentation given by H.M. Fischer. Microbiological Institute, ETH, Zürich, Switzerland, 10 Jan. 2013

Madigan et al., 2012

Gruber and Galloway, Nature, 2008

bacteroids in

infected plant cells

Selected rhizobial species and their legume hosts

Family	Genus	Species (examples)	Host plant(s)	
α-proteobacteria				
Rhizobiaceae	Rhizobium	<i>R. leguminosarum</i> bv. viciae	Pisum sativum (pea)	
		<i>R. leguminosarum</i> bv. trifolii	<i>Trifolium</i> (clover), <i>Vicia</i> (vetch)	
		R. etli	Phaseolus vulgaris (bean)	
	Sinorhizobium (Ensifer)	S. (E.) meliloti	Medicago sativa (alfalfa)	
Phyllobacteriaceae	Mesorhizobium	M. loti	Lotus japonicus	
Bradyrhizobiaceae	Bradyrhizobium	B. japonicum	Glycine max (soybean), mungbean, cowpea, siratro	
		<i>B. japonicum</i> ORS278 (photosynthetic, lacks <i>nodA</i> and <i>nodC</i>	Aeschynomene spp., (root and stem nodules)	
Methylobacteriacea	Methylobacterium	M. nodulans	Crotalaria spp.	
Kanthobacteraceae Azorhizobium		A. caulinodans (genuine diazotroph)	<i>Sesbania rostrata</i> (root and stem nodules)	
β-proteobacteria				
Burkholderiaceae	Burkholderia	B. phymatum	Mimosa spp., Phaseolus vulgaris	
	Cupriavidus	C. taiwanensis	Mimosa ssp.	

Source: Presentation given by H.M. Fischer. Microbiological Institute, ETH, Zürich, Switzerland, 10 Jan. 2013.

Rhizobia efficacy is strain dependent

Rhizobium strain	<i>Total N (mg plant⁻¹)</i>		
N8	3.7		
N5	3.4		
S 6	3.0		
S36	2.5		
C100	2.0		

Rhizobia inoculants Legumes seed-inoculation technology



Liquid inoculants





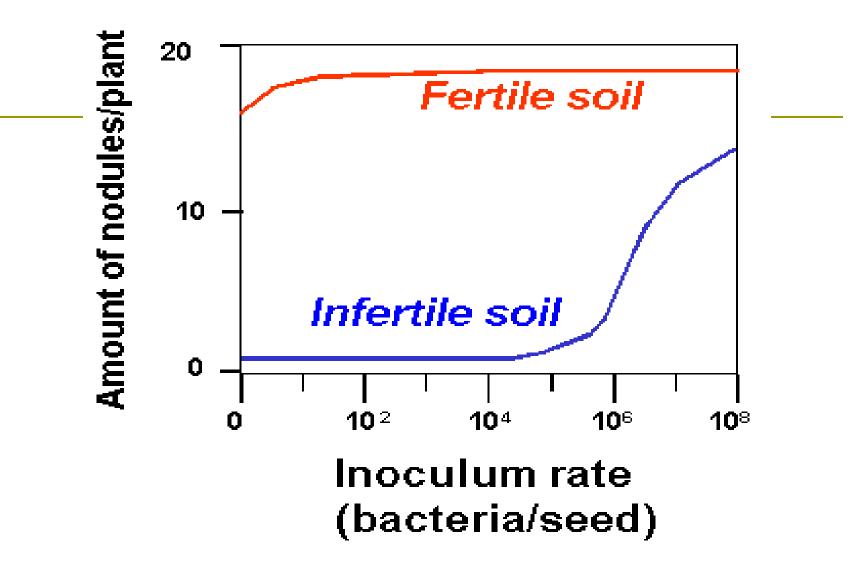
Peat based inoculants



Granular peat based inoculum







Soya grain yield after inoculation with Rhizobium & fertilizing with N

Rhizobium	N/ha	opbr_15%	relatief	eiwit%	N_zaad
coating	0	3797	100	37.0	207
coating	50	3619	95	35.9	192
coating	100	3583	94	35.4	188
coating	150	3428	90	35.3	176
zonder	0	1629	43	26.6	64
zonder	50	2130	56	26.8	84
zonder	100	2244	59	28.9	96
zonder	150	2593	68	30.2	116
lsd		288		1.2	18
		Kg/ha i	relativ	%	N in seed

PRAKTIJKONDERZOEK PLANT & OMGEVING WAGENINGEN UR

protein

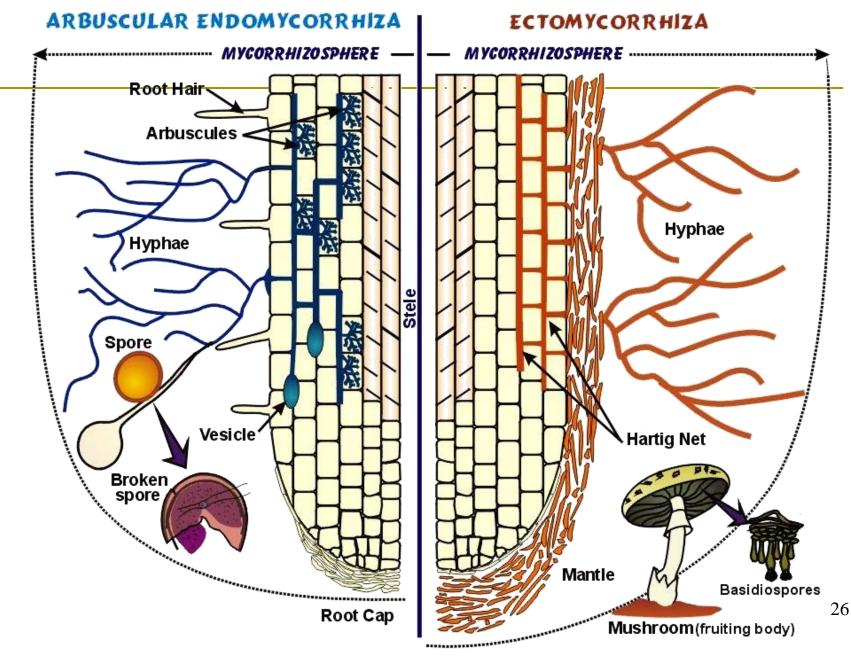
Practical value of Rhizobium-Legume symbiosis

- Effective legume-Rhizobium symbiosis can fix 100-400 kg / ha nitrogen (from air); corresponds to ca 220 – 900 kg Urea
- Fixed N₂ is used for own legume production. Surplus of N₂ for follow crop N₂ accumulation of 50-200 kg/ha for rotation crop possible
- Legume-Rhizobium symbiosis is sensitive to mineral nitrogen fertilization at planting; it is not recommended to add any Nitrogen when inoculating with Rhizobium
- Effective symbiosis needs phosphate (in fertile soils P is biologically provided by arbuscular mycorrhiza) & special micro-nutrients (Mo, Co, Fe)

Plant Root Symbionts:

Rhizobia for N2 fixation Mycorrhiza (for nutrient uptake and transport)

Mycorrhizas – mutualistic symbioses





Ectomycorrhizal fungi (about 5000): Most are Basidiomycetes; also some Ascomycetes (truffles); Zygomycetes (ectendo = Endogone)

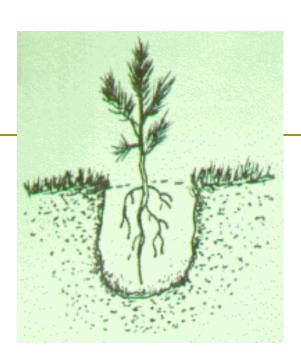


Lactarius (left) and Boletus (right)

About 2000 plant species form ectomycorrhiza, mainly trees: Pinaceae, Cupressaceae, Fagaceae, Betulaceae, Salicaceae, Dipterocarpoideae, Myrtaceae, also Eucalyptus species - All are of obligate mycotrophic character















Ectomycorhhizal inoculum applied in nursery, or at time of transplant

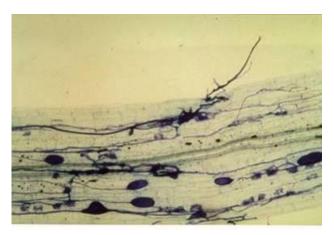




Genetic Resources of Rhizosphere Microorganism

Arbuscular Mycorrhiza =endomyorrhiza

Fungi belong to Glomeromycota Multinucleous organisms

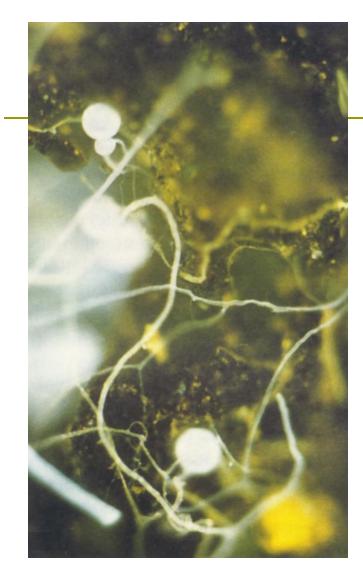


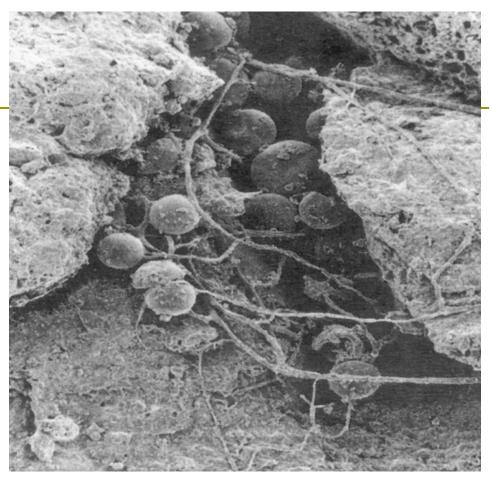


Formed by >70 % of plant species of plant kingdom with about 300 fungal species of the Glomeromycota.

Important crops like all cereals & grasses, potatoes, vegetables, flowers, fruit trees form mycorrhiza.

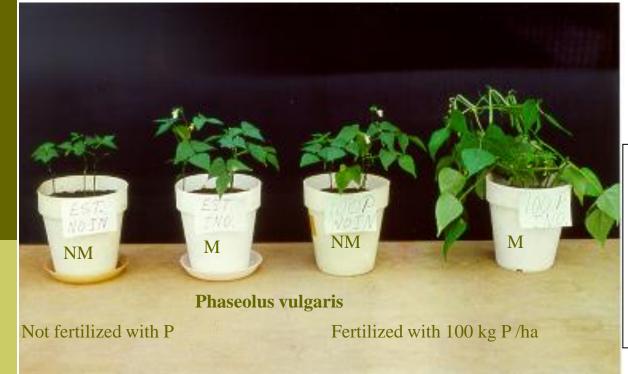
Some crops never form mycorrhiza: rape, sugarbeet, brassicas Some need special mycorrhiza: blue berries, Ericaceae, orquideae





Root external mycorrhizal mycelium: increase of exploited soil volume -6-200 times higher than by roots alone

Genetic Resources of Rhizosphere Microorganism



Mycorrhiza absorbs Plant available phosphate More efficient than the root

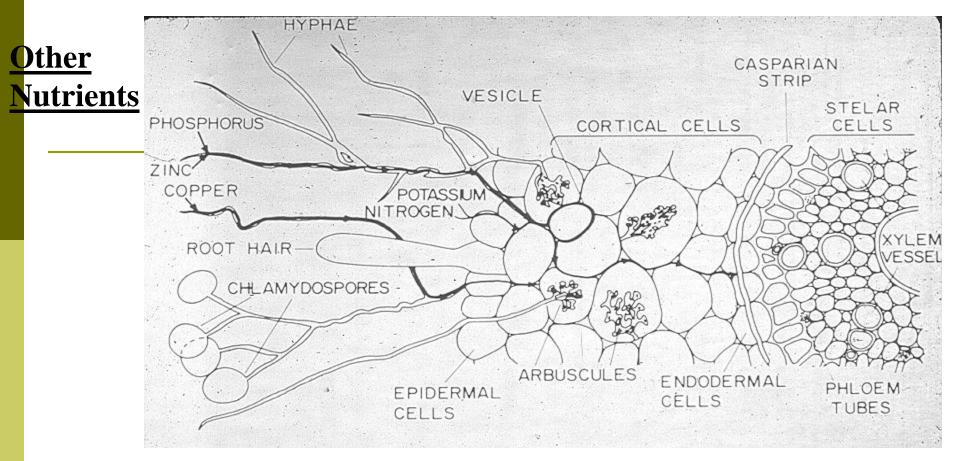
Phosphat is a growth limiting element in soils, and is mainly needed at the beginning of season

Dependency of plants on mycorrhiza: Some are obligate Some are facultative

Root Morphology dependent



Onion bulbs grown in soilless mixture (peat/vermiculite/perlite) and treated (A) or not (B) treated with BACTOLiVE AM BASIC inoculum (after 3 weeks of inoculation



Macro nutrients: direct uptake and transport by hyphae: P, NO₃⁻, NH₄⁺, Ca, Mg, (K? - via mass flow?) Micro nutrients: direct uptake & transport by hyphae: Zn, Cu, S, B, Mo (essential nutrient) Cd, Ni, Sr, Se (heavy metal) Br, J (not essential elements) Indirect? (Higher concentrations in mycorrhizal plants): Fe, Mn, Cl (assumed also Na, Real Stransport Rhizosphere Microorganism

Other benefits of Arbuscular Mycorrhiza (AM)

Plant water relations

Plant health (bio control of root pathogens and

more resistant to nematodes)

- Interaction with soil microorganisms
 - AM improve biological N2 fixation by Rhizobia
 - AM improve plant growth promoting bacteria and P solubilizers



Nematode control in vegetables by arbuscular mycorrhiza



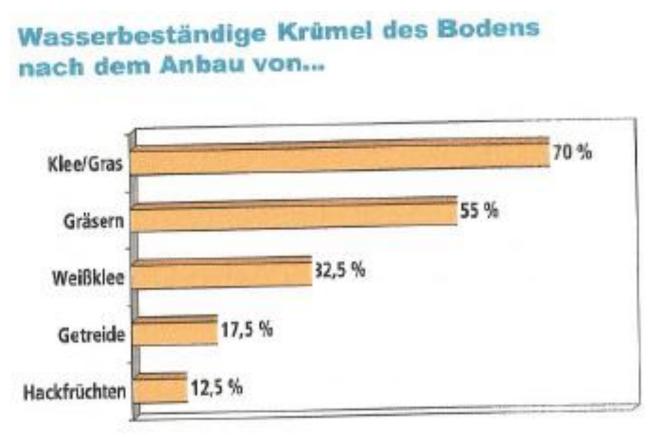
GLOMALIN

Very important part of SOIL FERTILITY

Glomalin glues soil particles together and forms stable soil aggregates

Potentially the most important benefit of mycorrhizal fungi for a sustainable agricultural production

Water stable aggregates (by Glomalin)



Grasses and legume are heavy mycorrhizal – excellent soil aggregation Sugar beet doesnot form mycorrhiza – no soil aggregation

WHERE WE WAS A PROPERTY OF THE PROPERTY OF THE

Water stable aggregates



Practical value of arbuscular mycorrhiza

- □ Savings of 50% Phosphate fertilizer, if mycorrhiza is promoted
- □ Making more effective use of fertilized P
- □ Making use of low soluble P-fertilizers (rock phosphates)
- More balanced uptake of fertilizer nutrients (N, K, Mg, micronutrients)= with better plant growth
- Better survival of plants during short drought periods
- Significant better N-fixation by legumes via Rhizobium
- Plays role in soil health (nematode control) and in soil aggregation (soil fertility)

Summary

- Without roots no shoots = without roots no yield
- Early root development depends (in general) on availability of phosphate; it is well known that too much nitrogen overproportionally favours shoot development
- Optimum root development at the start of the vegetation is important for yield (nutrient use efficacy, water use efficacy)
- It appears that a lot of research is still required what roots concern, and what their symbionts concern more needs to be done to make optimum use of the natural resource "root" in agricultural production (it appears that more emphasis is placed on "fertilizing" the above ground part of crops root research means: a lot of dirty work)
- Roots are generally living in association with surrounding microorganisms, or in symbiosis with them – without microorganisms there is no plant nutrition & likely low yields.

THANK YOU VERY MUCH